

REVIEW

Toward 'SMART' stem cells

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Stem cell research is at the heart of regenerative medicine, which holds great promise for the treatment of many devastating disorders. However, in addition to hurdles posed by well-publicized ethical issues, this emerging field presents many biological challenges. What is a stem cell? How are embryonic stem cells different from adult stem cells? What are the physiological bases for therapeutically acceptable stem cells? In this editorial review, I will briefly discuss these

superficially simple but actually rather complex issues that surround this fascinating cell type. The goal of this special issue on stem cells in Gene Therapy is to review some fundamental and critical aspects of current stem cell research that have translational potential.

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The promise of stem cell research

Stem cell research is moving to center stage in the biomedical field. The main driving force for this is the demand for regenerative medicine, which offers hope for the treatment of many devastating degenerative diseases such as diabetes, Parkinson's disease and Alzheimer's disease. The field has already demonstrated what is considered a classic example, that is, hematopoietic stem cell (HSC) transplantation, which has been used in the clinic to cure or treat hematopoietic failures, immune deficiencies, leukemias and certain other cancers. There is great potential for significant health gains in a much broader scope through the use of stem cell therapy in conjunction with other medical modalities. For instance, the first-line treatment of chronic myeloid leukemia (CML)—a well known HSC cancer with small molecule ABL kinase inhibitors has been a great success story of molecular medicine.¹ However, CML remains an incurable disease likely due to the inability of the current inhibitors to eradicate CML-initiating cells or the so-called 'leukemic stem cells'.² To cure CML, it will be necessary to target CML-leukemic stem cells specifically.

Stem cell research has been associated with gene therapy since its establishment. It continues to provide a complementary aspect to gene therapy because the self-renewing and differentiative properties of stem cells make them the ideal vehicle for therapeutic transgenes in their progeny.⁴ The complete cure of adenosine deaminase-deficient patients by HSC gene therapy represents a

remarkable success in the use of combinational stem cell and gene therapies to manage an inherited disorder.⁵ A recent study demonstrated that HSCs can carry a specific antigenic protein which, once the HSCs differentiate into T cells, will eradicate a tumor.⁶ This suggests a broad application of HSCs therapy in conjunction with immunotherapy and gene therapy for treating a broad spectrum of cancers.

Given the fact that tissue stem cells exist in many organs throughout an organism's lifetime, it seems likely that these cells are involved in the pathogenesis of many diseases, although this cannot be definitively studied until the phenotype of a specific tissue stem cell type is identified. For example, it is known that many types of human leukemia are directly or indirectly transformed at the HSC level.^{7,8} Furthermore, the current cancer stem cell or tumor stem cell (TSC) hypothesis postulates that self-renewing cells within a tumor are responsible for tumor growth and renewal.^{9,10} Increasing evidence supports the existence of TSCs in solid tumors derived from many organs, including the human breast, brain, colon, pancreas and prostate.^{11–16} Stem cell culture toward a specific path or a particular disease state may resemble normal developmental process or pathogenesis, thus offering a new platform for drug discovery and pharmacology/toxicology studies.¹⁷

For those researchers who are not directly involved in stem cell research, it is worth mentioning that embryonic stem cells (ESCs) have been used as a powerful and indispensable cellular vehicle to carry an exogenous transgene or to target an endogenous gene product in a specific tissue lineage or a differentiated cell type to study the function of a gene at different developmental stages. The discovery concerning the principles for gene targeting in mice via the use of ESCs by Mario Capecchi, Martin Evans and Oliver Smithies has recently won the Nobel Prize in Physiology or Medicine of 2007.

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About the concept of stem cells

Enormous advances in stem cell research have been made, especially since the first human ESC line was established.¹⁸ The stem cell research community is becoming a united scientific entity across basic science, clinical medicine, ethics and bioindustry, yet many concepts in the field remain poorly defined. Even the popular term 'stem cells' has no universally accepted definition.^{19–23} This ambiguity has caused confusion regarding what is being studied, especially among the general public.

The term 'stem cells' first appeared in the research literature at least 140 years ago.²⁶ It was originally used by embryologists to describe germline cells, and by hematologists to describe blood-forming cells.²⁶ In 1961, Till and McCulloch²⁷ established the spleen colony assay to define the mouse HSC with its ability to self-renew. Since then, the potential for differentiation and self-renewal has been considered to be two classical fundamental properties of a stem cell. However, these two properties must be retrospectively demonstrated, which is never an easy task in research.

The field of stem cell research has been significantly shaped by the isolation of mouse HSCs using flow cytometry, a process that can identify a definitive immunophenotype.²⁸ However, the specific immunophenotype of a stem cell is not fixed and it could be a moving target depending on many factors such as age, strain, species and host conditioning.^{29,30} (H Shen and T Cheng, unpublished data). Even for the 'best defined' mouse HSC, there is no single immunophenotype that has been used by everyone in the field. There is currently an emphasis on functional approaches to stem cell identification such as Hoechst efflux,^{31–34} however, these approaches have not been able to offer a definitive solution.³⁵ In general, functional and phenotypic characteristics are both important in defining a tissue stem cell, but the two are not always associated with each other. Unfortunately, stem cell rarity in the body precludes us from identifying stem cell types by the use of many conventional molecular analyses. Therefore, unlike many other tissue cell types, the definition of stem cell types at cellular and molecular levels is often nonoptimal and could be considered arbitrary in many studies. The variability that exists within what is defined as a tissue stem cell type of any particular organ may be largely responsible for discrepancies between published data regarding specific stem cell types. Therefore, one of the most important subjects for stem cell research is the continuing effort to define stem cell identity.

Embryonic stem cells versus adult stem cells

Perhaps the greatest cause of confusion for the general public—and even in some researchers' minds—is the use of the term 'stem cells' to describe both ESCs and tissue or adult stem cells (ASCs), two major general stem cell types. (For the purposes of this paper, I will use ASCs to represent all somatic tissue stem cells except ESCs, and therefore ASCs do not necessarily refer to stem cells from adults.) Although these stem cell types differ in many fundamental ways, they are often regarded as a similar

Table 1 Fundamental differences between ESC and ASC

	ESC	ASC
Origin	Blastocyst	Developed tissues
Proliferation	Indefinite	Limited
<i>in vitro</i> Differentiation spectrum	All the tissue types	Limited
Homing ability	No	Yes
Reconstitution efficiency	Low	High
Tumorigenesis	Teratoma	No or rare
Availability	Restricted (human)	Less restricted
Ethical issues	Severe (human subjects or human-xeno models)	Less
Clinical proof	No	Yes (HSC)
The most challenging technique in therapeutics	Efficient induction of specific tissue types without tumorigenesis	<i>In vitro</i> expansion without loss of physiological properties

Abbreviations: ASC, adult stem cell; ESC, embryonic stem cell; HSC, hematopoietic stem cell.

cell type. This misconception not only causes biological inaccuracies in describing these cells but also generates problems in the ethical debates over stem cell research. Given this situation, I would like to reiterate the key differences between ESCs and ASCs as illustrated in Table 1. In short, ESCs are cultured cell lines and an ESC equivalent has yet to be discovered *in vivo*. In contrast, ASCs are more physiologically relevant and thus can be readily used for autologous transplant and if a specific immune barrier can be managed—in the setting of allo-transplant.

It should be emphasized that the comparison in Table 1 does not suggest a superior importance of one stem cell type over another. Generally speaking, both are equally important. The choice of ESCs versus ASCs in therapeutics is dependent upon the specific type of diseased tissue that is receiving therapy. For some tissues, such as the pancreas in which ASCs have not been shown to be essential in tissue regeneration,³⁶ ESCs may be more valuable as a source of making functional β cells. In contrast, HSCs are known to be fully responsible for regenerating all of the blood cell lineages. If the current hurdle of HSC expansion can be overcome, we may not have to use ESC-derived HSCs for HSC therapy in some clinical circumstances such as autologous HSC transplant.

In addition to the individual uses to which ESCs and ASCs can be put, research into each of these types is critical to understanding the other. ESCs can be precursor cells for ASCs, and this process offers a useful model to study how ASCs are generated. As a result, the 'tricks' for expanding ASCs may be uncovered by studying ESC biology. On the other hand, understanding how ASCs operate physiologically is pivotal for the development into strategies for coaxing ESCs toward differentiation into therapeutically acceptable cells.

Critical aspects on the physiology of stem cells

Our ability to manipulate stem cells for therapeutic purposes is highly dependent on our understanding of

the mechanisms that underlie stem cell kinetics *in vivo*. Stem cells are defined by their capacity for self-renewal and differentiation, but there are only two multiple cellular functions that are critical for maintaining the homeostasis of ASCs *in vivo*.

Self-renewal

It is a common misconception that all stem cell self-renewal occurs in the same way that general cells proliferate. In fact, stem cells show two different methods of self-renewing: one is asymmetrical (one daughter stem cell and one differentiated cell after division) and the other is symmetrical (two daughter stem cells after division). ESCs can only undergo symmetrical self-renewing division, whereas ASCs (for example, HSCs) and neural stem cells (NSCs) are thought to undergo asymmetrical self-renewing division under homeostatic conditions.³⁷ At the cell population level, stem cells self-replicate at a probability between 0 and 1.0.²⁰ Thus, stem cell maintenance requires probability of 0.5 and a probability of less or more than 0.5 can result in exhaustion or expansion, respectively. The acceleration of HSC proliferation often results in the loss of HSC self-renewal potential after birth.³⁸⁻⁴⁰ The molecular basis for this cellular phenomenon has been shown in mice deficient in several cell cycle regulators such as p21 (Cip1/Waf1), Gfi-1 and Pten.³⁹⁻⁴¹

Because ESCs and ASCs self-renew in different manners, it is unlikely that the same molecular circuitry governs the self-renewal process in both cell types. It has been shown that transcription programs dictated by Oct4, Nanog and Sox2 play essential roles in ESC self-renewal.⁴² In ASCs, the mechanisms appear to be much more complex, although Bmi-1 has been shown to be shared between some ASC types, including HSCs, NSCs and mammary stem cells.⁴³⁻⁴⁶ Interestingly, this mechanism is also shared by some TSCs.⁴⁴ As a cell cycle inhibitor, p18 has been shown to play a unique role in the self-renewal of several stem cell types, including HSCs, lung stem cells and possibly NSCs.^{47,48} While self-renewal enables stem cells to continue even beyond the lifetime of the organisms from which they are harvested,⁴⁹ this ability appears to decrease with increasing age of the cell.⁵⁰ Ataxia telangiectasia mutated and p16 appear to play important roles in this decrease.⁵¹⁻⁵³

Maturation (differentiation)

Differentiation is the most direct cellular basis for using stem cells in regenerative medicine. All stem cells are able to differentiate into specified cell types under specific conditions. The spectrum of differentiation depends upon the stem cell type and the microenvironmental cues it receives. Unlike ESCs, which can differentiate into virtually any kind of cell in the body, the plasticity of ASCs toward different tissue types appears to be limited,⁵⁴⁻⁵⁶ although this issue continues to be debatable depending upon the identity of actual input 'stem cells' in a given study.^{57,58}

The molecular basis for the choice between self-renewal and differentiation is the central question in stem cell biology. There have been two general theories regarding the mechanisms: instructive (environment-dependent) and stochastic (environment-independent).^{19,59} As mentioned above, unlike ESCs in which

differentiation and self-renewal are uncoupled, ASCs can undergo asymmetrical division resulting in concurrence of self-renewal and differentiation.^{60,61} As largely demonstrated in hematopoiesis, differentiation is a sequential process that ultimately yields fully mature cell populations via hematopoietic progenitor cells,⁶² although this was challenged by an alternative model in which a specific cell cycle position, rather than a hierarchical position in the differentiation cascade, determines whether a primitive cell functions as a stem or a progenitor cell.⁶³ According to the hierarchical model, however, stem cell number is not the sole determinant for the size of the final mature cells. In fact, because intermediate progenitor cells directly yield mature cell populations, self-regeneration of these progenitor cells must also be crucial or even more important in producing mature cells under homeostatic conditions.²⁵ Therefore, factors regulating stem cell repopulation efficiency (meaning the function per stem cell) are important but have been less emphasized, although this paradigm was partially illustrated in the absence of p27.⁶⁴

Apoptosis (cell death)

In theory, apoptosis is also a fate choice of stem cells. But under homeostatic conditions, in which stem cells undergo asymmetrical divisions to maintain a constant number of stem cells, apoptosis may be a less frequent event. However, once symmetrical self-renewing division occurs and the stem cell pool size is increased, apoptosis is likely one of the mechanisms used to maintain the stem cell pool at its proper size. Stem cells must have the capability to die in order to prevent the overgrowth of stem cells that may lead to cancer.

In general, stem cell apoptosis is a relatively poorly studied area. Although the antiapoptotic protein Bcl-2 has been shown to increase HSC number in a transgenic model,⁶⁵ whether it acts directly via suppression of an apoptosis mechanism remains to be further defined, given the fact that Bcl-2 could also alter HSC cycling.⁶⁵ While the p53 pathway has been shown to be involved in stem cell senescence,^{66,67} its direct role in stem cell apoptosis has not been clearly defined.

Resting mode (quiescence)

Adult stem cells have been thought to be resistant to many physiological stimuli and pathophysiological insults due to their ability to maintain themselves for the lifetime of an organism. It is known that HSCs do not respond to many hematopoietic growth factors that affect hematopoietic progenitor cell populations under physiological conditions.⁶⁸⁻⁷⁰ As a result, HSCs are relatively quiescent in cell cycle. Based on this characteristic, suicide approaches have been used to spare stem cells from their progeny when progenitor-specific growth factors and the S-phase-specific toxin 5-FU were sequentially applied in culture.⁷¹⁻⁷³ Therefore, the resting mode in which the majority of ASCs are maintained under homeostasis *in vivo* appears to be an important self-protective mechanism of the cells. However, the molecular basis for this cellular defense mechanism is largely unclear. While a number of potential molecular players such as p21, Gfi-1, MEF/ELF4 and angiotensin signaling have been reported,^{39,40,46,74} a common unifying mechanism has yet to be defined. In addition, potential

links of stem cell quiescence to other important cellular defense mechanisms, such as multidrug resistance and DNA repair possessed by stem cells,⁷⁵⁻⁷⁷ are important areas that need to be investigated.

Trafficking (migration)

The ability of stem cells to traffic or migrate is best described with HSCs. HSCs have an active motility.^{78,79} They constantly move in and out of their bone marrow niche and even circulate in the blood under homeostatic conditions.^{80,81} Similarly, physiological stem cells are able to travel to their niche in a specific organ. This ability appears to be gradually acquired during embryonic development⁸² and is a prerequisite for stem cells to reach and reside in damaged tissues in therapeutic scenarios. Therefore, the success of an HSC transplant is largely dependent upon the homing efficiency of transplanted HSCs in the marrow. Specific manipulations of homing molecules may enhance HSC engraftment efficiency upon transplantation. Such manipulation was exemplified in a recent study which demonstrated that targeting CD26 can significantly enhance HSC therapy efficiency in both mouse and human hematopoietic systems.^{83,84}

Together, these five minimal functional states of stem cells (self-renewal, maturation, apoptosis, resting mode and trafficking) constitute an interesting 'SMART' model for maintaining stem cell homeostasis *in vivo* (Figure 1). In fact, critical roles of the multicellular niche in stem cell maintenance further reinforce the physiological definition of ASCs *in vivo* as illustrated by these functional features.^{85,86} The lack of any of these 'SMART' features would make stem cells much less physiological and particularly useless in therapeutics. Given these highly complex and controlled features *in vivo*, the great variability of stem cell phenotypes might be largely due to specific functional states of given stem cells along these different fate choices. According to this model (that is certainly suitable for some well-defined ASCs such as HSCs), ESCs are apparently a more homogeneous but defective stem cell population because they can only self-renew in a symmetrical manner. ESCs do not have the ability to travel purposefully or migrate to an injured site and exert minimal or uncoupled functions of other

features (Table 1). Even in the scenario where a differentiated cell type can be reprogrammed to an ES-like cell type^{87,88} or ES equivalents can be generated via epiblasts,⁸⁹ these stem cell lines will have the same shortcomings as ESCs.

Unfortunately, however, TSCs also have these 'SMART' features and, in fact, TSCs must be able to outcompete their normal counterpart when they become a dominant phenotype. Regarding therapeutic manipulations, especially those involving genetic approaches, a fundamental challenge is to produce therapeutically acceptable stem cell products while avoiding the production of tumorigenic stem cells. For example, while overexpression of HoxB4 has been shown to be very effective in HSC expansion and leads to no increase of leukemogenesis in mice,^{90,91} a recent study reported that dogs transplanted with HoxB4-expanded HSCs were susceptible to leukemia development.⁹² Therefore, defining the molecular boundary between normal ASC and TSC is vital for the safe expansion of ASCs as well as for the development of methods to selectively target TSCs for therapeutic purposes.⁹³

Final notes

Stem cell research shows promise for the treatment and/or cure of many devastating diseases for which we do not have effective treatments at the present time. Moreover, stem cell research provides new opportunities to increase the effectiveness of existing medical modalities. Our ability to manipulate stem cells for therapeutic purposes is directly dependent upon understanding the biology of this fascinating cell type. However, stem cell biology is still in its immature stage and thus has an enormous potential to grow. We are facing many critical challenges in its biological research as well as ethics. The 'SMART' model illustrated in this review (Figure 1) is not intended to offer a new definition for stem cells but rather to exemplify the physiological relevance of stem cells for their potential therapeutic applications. It should also be emphasized that stem cell therapy cannot stand by itself and combinational therapies, such as a combined stem cell and gene therapy, are likely the common scenarios for future clinical practice in this area.

In this special issue of *Gene Therapy*, which focuses on stem cells, limited space prevents us from covering all aspects of the field. Rather, we intend to showcase important topics that we feel have been relatively underemphasized in the literature. These topics are relevant but not limited to gene therapy, and all are directly or indirectly related to potential stem cell therapeutic development.

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Figure 1 The 'SMART' physiological features of stem cells *in vivo*.

References

- 1 Drucker BJ. Inhibition of the Bcr-Abl tyrosine kinase as a therapeutic strategy for CML. *Oncogene* 2002; 21: 8541-8546.
- 2 Ren R. Mechanisms of BCR-ABL in the pathogenesis of chronic myelogenous leukaemia. *Nat Rev Cancer* 2005; 5: 172-183.
- 3 Hu Y, Swerdlow S, Duffy TM, Weinmann R, Lee FY, Li S. Targeting multiple kinase pathways in leukemic progenitors and stem cells is essential for improved treatment of *Ph1* leukemia in mice. *Proc Natl Acad Sci USA* 2006; 103: 16870-16875.
- 4 Bordignon C, Roncarolo MG. Therapeutic applications for hematopoietic stem cell gene transfer. *Nat Immunol* 2002; 3: 318-321.
- 5 Booth C, Hershfield M, Notarangelo L, Buckley R, Hoenig M, Mahlaoui N *et al.* Management options for adenosine deaminase deficiency; proceedings of the EBMT satellite workshop (Hamburg, March 2006). *Clin Immunol* 2007; 123: 139-147.
- 6 Yang L, Baltimore D. Long-term *in vivo* provision of antigen-specific T cell immunity by programming hematopoietic stem cells. *Proc Natl Acad Sci USA* 2005; 102: 4518-4523.
- 7 Bonnet D, Dick JE. Human acute myeloid leukemia is organized as a hierarchy that originates from a primitive hematopoietic cell. *Nat Med* 1997; 3: 730-737.
- 8 Jamieson CH, Allles LE, Dylla SJ, Muijters M, Jones C, Zehnder JL *et al.* Granulocyte-macrophage progenitors as candidate leukemic stem cells in blast-crisis CML. *Engl J Med* 2004; 351: 657-667.
- 9 Reya T, Morrison SJ, Clarke MF, Weissman IL. Stem cells, cancer, and cancer stem cells. *Nature* 2001; 414: 105-111.
- 10 Clarke MF, Dick JE, Dirks PB, Evans CJ, Jamieson CH, Jones DL *et al.* Cancer stem cells—perspectives on current status and future directions: AACR Workshop on Cancer Stem Cells. *Cancer Res* 2006; 66: 9339-9344.
- 11 Al-Hajj M, Wicha MS, Benito-Hernandez A, Morrison SJ, Clarke MF. Prospective identification of tumorigenic breast cancer cells. *Proc Natl Acad Sci USA* 2003; 100: 3983-3988.
- 12 Singh SK, Clarke ID, Terasaki M, Bonn VE, Hawkins C, Squire J *et al.* Identification of a cancer stem cell in human brain tumors. *Cancer Res* 2003; 63: 5821-5828.
- 13 Hermstad HD, Nakano I, Lazareff JA, Masterman-Smith M, Geschwind DH, Brunker-Fraser M *et al.* Cancerous stem cells can arise from pediatric brain tumors. *Proc Natl Acad Sci USA* 2003; 100: 15178-15183.
- 14 Li C, Heidt DG, Dalerba P, Burant CF, Zhang L, Adsay V *et al.* Identification of pancreatic cancer stem cells. *Cancer Res* 2007; 67: 1030-1037.
- 15 O'Brien CA, Pollett A, Gallinger S, Dick JE. A human colon cancer cell capable of initiating tumor growth in immunodeficient mice. *Nature* 2007; 445: 106-110.
- 16 Collins AT, Berry PA, Hyde C, Stower MJ, Maitland NJ. Prospective identification of tumorigenic prostate cancer stem cells. *Cancer Res* 2005; 65: 10946-10951.
- 17 Pouton CW, Haynes JM. Embryonic stem cells as a source of models for drug discovery. *Nat Rev Drug Discov* 2007; 6: 605-616.
- 18 Thomson JA, Itskovitz-Eldor J, Shapiro SS, Waknitz MA, Swiergiel JJ, Marshall VS *et al.* Embryonic stem cell lines derived from human blastocysts. *Science* 1998; 282: 1145-1147.
- 19 Metcalf D. On hematopoietic stem cell fate. *Immunity* 2007; 26: 669-673.
- 20 Potten MLaC (ed). *Stem Cells and Cellular Pedigrees—A Conceptual Introduction*. Academic Press: London, 1997, 1pp.
- 21 Blau HM, Brazelton TR, Weinmann JM. The evolving concept of a stem cell: entity or function? *Cell* 2001; 105: 829-841.
- 22 Zippori D. The nature of stem cells: state rather than entity. *Nat Rev Genet* 2004; 5: 873-878.
- 23 Parker GC, Anastasova-Kristeva M, Broxmeyer HE, Dodge WH, Eisenberg LM, Gehring UM *et al.* Stem cells: shibboleths of development. *Stem Cells Dev* 2004; 13: 579-584.
- 24 Parker GC, Anastasova-Kristeva M, Eisenberg LM, Rao M, Williams MA, Sanberg PR *et al.* Stem cells: shibboleths of

- development, part II: toward a functional definition. *Stem Cells Dev* 2005; 14: 463-469.
- 25 Metcalf D. Concise review: hematopoietic stem cells and tissue stem cells: current concepts and unanswered questions. *Stem Cells* 2007; 10: 2390-2395.
- 26 Ramalho-Santos M, Willenbring H. On the origin of the term 'stem cell'. *Cell Stem Cell* 2007; 1: 35-38.
- 27 Till JE, McCulloch CE. A direct measurement of the radiation sensitivity of normal mouse bone marrow cells. *Radiat Res* 1961; 14: 213-222.
- 28 Spangrude GJ, Heimfeld S, Weissman IL. Purification and characterization of mouse hematopoietic stem cells [published erratum appears in *Science* 1989 Jun 22;444(908):1030]. *Science* 1988; 241: 58-62.
- 29 Spangrude GJ, Brooks DM, Tumas DB. Long-term repopulation of irradiated mice with limiting numbers of purified hematopoietic stem cells: *in vivo* expansion of stem cell phenotype but not function. *Blood* 1995; 85: 1006-1016.
- 30 Spangrude GJ, Brooks DM. Mouse strain variability in the expression of the hematopoietic stem cell antigen Ly-Ga/E by bone marrow cells. *Blood* 1993; 82: 3327-3332.
- 31 Goodell MA, Brose K, Paradis G, Conner AS, Mulligan RC. Isolation and functional properties of murine hematopoietic stem cells that are replicating *in vivo*. *J Exp Med* 1996; 183: 1797-1806.
- 32 Jones RJ, Wagner JE, Celano P, Zicha MS, Sharkey SJ. Separation of pluripotent haematopoietic stem cells from spleen colony-forming cells [see comments]. *Nature* 1990; 347: 188-189.
- 33 Juopperi TA, Schuler W, Yuan X, Collector MJ, Dang CV, Sharkey SJ. Isolation of bone marrow-derived stem cells using density-gradient separation. *Exp Hematol* 2007; 35: 335-341.
- 34 Hess DA, Meyerrose TE, Wirhlin L, Craft TP, Herrlich PE, Creer MH *et al.* Functional characterization of highly purified human hematopoietic repopulating cells isolated according to aldehyde dehydrogenase activity. *Blood* 2004; 104: 1648-1655.
- 35 Morita Y, Ena H, Yamazaki S, Nakachi H. Non-side-population hematopoietic stem cells in mouse bone marrow. *Blood* 2006; 108: 2850-2856.
- 36 Dor Y, Brown J, Martinez OL, Melton DA. Adult pancreatic beta-cells are formed by self-duplication rather than stem-cell differentiation. *Nature* 2004; 429: 41-46.
- 37 Morrison SJ, Kimble J. Asymmetric and symmetric stem-cell divisions in development and cancer. *Nature* 2006; 441: 1068-1074.
- 38 Bowie MB, McKnight KD, Kent DG, McCaffrey L, Hoodless PA, Eaves CJ. Hematopoietic stem cells proliferate until birth and show a reversible phase-specific engraftment defect. *J Clin Invest* 2006; 116: 2808-2816.
- 39 Cheng T, Rodriguez N, Shen H, Yang Y, Dombkowski D, Sykes M *et al.* Hematopoietic stem cell quiescence maintained by p21(cip1/waf1). *Science* 2000; 287: 1804-1808.
- 40 Hock H, Hamblin MJ, Rooke HM, Schindler JW, Saleque S, Fujiwara Y *et al.* Gfi-1 restricts proliferation and preserves functional integrity of hematopoietic stem cells. *Nature* 2004; 431: 1002-1007.
- 41 Zhang J, Grindley JC, Yin T, Jayasinghe S, He XC, Ross JT *et al.* PTEN maintains hematopoietic stem cells and acts in lineage choice and leukaemia prevention. *Nature* 2006; 441: 518-522.
- 42 Cavaleri F, Scholer HR. Nanog: a new recruit to the embryonic stem cell orchestra. *Cell* 2003; 113: 551-552.
- 43 Park IK, Qian D, Kiel M, Becker MW, Pihalja M, Weissman IL *et al.* Bmi-1 is required for maintenance of adult self-renewing hematopoietic stem cells. *Nature* 2003; 423: 302-305.
- 44 Lessard J, Sauvageau G. Bmi-1 determines the proliferative capacity of normal and leukemic stem cells. *Nature* 2003; 423: 255-260.

- 45 Molofsky AV, Pardi R, Iwashita T, Park IK, Clarke MF, Morrison SJ. Bmi-1 dependence distinguishes neural stem cell self-renewal from progenitor proliferation. *Nature* 2003; 425: 962-967.
- 46 Lacrazza HD, Yamada T, Liu Y, Miyata Y, Sivina M, Nunes J *et al.* The transcription factor MER/ELF4 regulates the quiescence of primitive hematopoietic cells. *Cancer Cell* 2006; 9: 175-187.
- 47 Yuan Y, Shen H, Franklin DS, Scadden DT, Cheng T. *In vivo* self-renewing divisions of hematopoietic stem cells are increased in the absence of the early G1-phase inhibitor, p18INK4C. *Nat Cell Biol* 2004; 6: 436-442.
- 48 Pei XH, Bai F, Smith MD, Xiong Y. p18INK4C collaborates with Men1 to constrain lung stem cell expansion and suppress non-small-cell lung cancers. *Cancer Res* 2007; 67: 3162-3170.
- 49 Yu H, Yuan Y, Shen H, Cheng T. Hematopoietic stem cell exhaustion impacted by p18INK4C and p21Cip1/Waf1 in opposite manners. *Blood* 2006; 107: 1200-1206.
- 50 Geiger H, Van Zant G. The aging of lympho-hematopoietic stem cells. *Nat Immunol* 2002; 3: 329-333.
- 51 Ito K, Hirose A, Arai F, Matsuoaka S, Takubo K, Hamaguchi I *et al.* Regulation of oxidative stress by ATM is required for self-renewal of hematopoietic stem cells. *Nature* 2004; 431: 997-1002.
- 52 Molofsky AV, Sluskey SG, Joseph NM, He S, Pardi R, Krishnamurthy J *et al.* Increasing p16(INK4a) expression decreases forebrain progenitors and neurogenesis during aging. *Nature* 2006; 443: 448-452.
- 53 Janzen V, Forkert R, Fleming HE, Saito Y, Waring MT, Dombkowski DM *et al.* Stem-cell ageing modified by the cyclin-dependent kinase inhibitor p16(INK4a). *Nature* 2006; 443: 421-426.
- 54 Wagers AJ, Sherwood RJ, Christensen JL, Weissman IL. Little evidence for developmental plasticity of adult hematopoietic stem cells. *Science* 2002; 297: 2256-2259.
- 55 Wagers AJ, Weissman IL. Plasticity of adult stem cells. *Cell* 2004; 116: 639-648.
- 56 Sung LY, Gao S, Shen H, Yu H, Song Y, Smith SL *et al.* Differentiated cells are more efficient than adult stem cells for cloning by somatic cell nuclear transfer. *Nat Genet* 2006; 38: 1323-1328.
- 57 Jang YY, Sharikis SJ. Stem cell plasticity: a rare cell, not a rare event. *Stem Cell Rev* 2005; 11: 45-51.
- 58 Theise ND, Krause DS, Sharikis S. Comment on 'Little evidence for developmental plasticity of adult hematopoietic stem cells'. *Science* 2003; 299: 1317; author reply 1317.
- 59 Metcalf D. Lineage commitment and maturation in hematopoietic cells: the case for extrinsic regulation. *Blood* 1998; 92: 345-347; discussion 352.
- 60 Altar EC, Scadden DT. Regulation of hematopoietic stem cell growth. *Leukemia* 2004; 18: 1760-1768.
- 61 Molofsky AV, Pardi R, Morrison SJ. Diverse mechanisms regulate stem cell self-renewal. *Curr Opin Cell Biol* 2004; 16: 700-707.
- 62 Morrison SJ, Uchida N, Weissman IL. The biology of hematopoietic stem cells. *Annu Rev Cell Dev Biol* 1995; 11: 35-71.
- 63 Quesenberry PJ, Colvin GA, Lambert JF. The chiascuro stem cell: a unified stem cell theory. *Blood* 2002; 100: 4266-4271.
- 64 Cheng T, Rodrigues N, Dombkowski D, Sier S, Scadden D. Stem cell repopulation efficiency but not pool size is governed by p27. *Nat Med* 2000; 6: 1235-1240.
- 65 Domen J, Cheshier SH, Weissman IL. The role of apoptosis in the regulation of hematopoietic stem cells: overexpression of Bcl-2 increases both their number and repopulation potential. *J Exp Med* 2000; 191: 253-264.
- 66 TeKippe M, Harrison DE, Chen J. Expansion of hematopoietic stem cell phenotype and activity in Trp53-null mice. *Exp Hematol* 2003; 31: 521-527.
- 67 Dumble M, Moore L, Chambers SM, Geiger H, Van Zant G, Goodell MA *et al.* The impact of altered p53 dosage on hematopoietic stem cell dynamics during aging. *Blood* 2007; 109: 1736-1742.
- 68 Cheng T, Scadden DT. Cell cycle entry of hematopoietic stem and progenitor cells controlled by distinct cyclin-dependent kinase inhibitors. *Int J Hematol* 2002; 75: 460-465.
- 69 Bradford GB, Williams B, Rossi R, Bertoncello I. Quiescence, cycling, and turnover in the primitive hematopoietic stem cell compartment. *Exp Hematol* 1997; 25: 445-453.
- 70 Gother A, Pyatt R, McMahon J, Rice S, Srouf EF. Functional heterogeneity of human CD34(+) cells isolated in subcompartments of the G0/G1 phase of the cell cycle. *Blood* 1997; 90: 4384-4393.
- 71 Berardi AC, Wang A, Levine JD, Lopez P, Scadden DT. Functional isolation and characterization of human hematopoietic stem cells. *Science* 1995; 267: 104-108.
- 72 Lerner C, Harrison DE. 5-Fluorouracil spares hematopoietic stem cells responsible for long-term repopulation. *Exp Hematol* 1990; 18: 114-118.
- 73 Bertolini F, Battaglia M, Soligo D, Corsini C, Curioli C, Lazzari L *et al.* Stem cell candidates purified by liquid culture in the presence of steel factor, IL-3, and 5FU are strictly stroma-dependent and have myeloid, lymphoid, and megakaryocytic potential. *Exp Hematol* 1997; 25: 350-356.
- 74 Arai F, Hirose A, Ohnura M, Sato H, Matsuoaka S, Takubo K *et al.* Tie2/Angiopoietin-1 signaling regulates hematopoietic stem cell quiescence in the bone marrow niche. *Cell* 2004; 118: 149-161.
- 75 Chaudhary PM, Roninson IB. Expression and activity of P-glycoprotein, a multidrug efflux pump, in human hematopoietic stem cells. *Cell* 1991; 65: 85-94.
- 76 Rossi DJ, Seita J, Czechowicz A, Bhattacharya D, Bryder D, Weissman IL. Hematopoietic stem cell quiescence attenuates DNA damage response and permits DNA damage accumulation during aging. *Cell Cycle* 2007; 6: 2371-2376.
- 77 Rossi DJ, Bryder D, Seita J, Nussenzweig A, Hoeijmakers J, Weissman IL. Deficiencies in DNA damage repair limit the function of hematopoietic stem cells with age. *Nature* 2007; 447: 725-729.
- 78 Frimberger AE, McAuliffe CI, Werne KA, Tuft RA, Fogarty KE, Benoit BO *et al.* The fleet feet of hematopoietic stem cells: rapid motility, interaction and proteolysis. *Br J Haematol* 2001; 112: 644-654.
- 79 Frimberger AE, Sterling AJ, Quesenberry PJ. An *in vitro* model of hematopoietic stem cell homing demonstrates rapid homing and maintenance of engraftable stem cells. *Blood* 2001; 98: 1012-1018.
- 80 Lapidot T, Dar A, Kollet O. How do stem cells find their way home? *Blood* 2005; 106: 1901-1910.
- 81 Wright DE, Bowman EP, Wagers AJ, Butcher EC, Weissman IL. Hematopoietic stem cells are uniquely selective in their migratory response to chemokines. *J Exp Med* 2002; 195: 1145-1154.
- 82 Lee BC, Cheng T, Adams GB, Altar EC, Miura N, Lee SB *et al.* P2Y-like receptor, GPR105 (P2Y14), identifies and mediates chemotaxis of bone-marrow hematopoietic stem cells. *Genes Dev* 2003; 17: 1592-1604.
- 83 Christopherson II KW, Hangoc G, Mantel CR, Broxmeyer HE. Modulation of hematopoietic stem cell homing and engraftment by CD26. *Science* 2004; 305: 1000-1003.
- 84 Campbell TB, Hangoc G, Liu Y, Pollok K, Broxmeyer HE. Inhibition of CD26 in human cord blood CD34+ cells enhances their engraftment of nonobese diabetic/severe combined immunodeficiency mice. *Stem Cells Dev* 2007; 16: 347-354.
- 85 Scadden DT. The stem-cell niche as an entity of action. *Nature* 2006; 441: 1075-1079.
- 86 Fuchs E, Tumber T, Guasch G. Socializing with the neighbors: stem cells and their niche. *Cell* 2004; 116: 769-778.

- 87 Takahashi K, Yamanaka S. Induction of pluripotent stem cells from mouse embryonic and adult fibroblast cultures by defined factors. *Cell* 2006; 126: 663-676.
- 88 Meissner A, Wernig M, Jaenisch R. Direct reprogramming of genetically unmodified fibroblasts into pluripotent stem cells. *Nat Biotechnol* 2007; 25: 1177-1181.
- 89 Tesar PJ, Chenoweth JG, Brook FA, Davies TJ, Evans EP, Mack DL, et al. New cell lines from mouse epiblast share defining features with human embryonic stem cells. *Nature* 2007; 448: 196-199.
- 90 Sauvageau G, Thorsteinsdottir U, Faves CJ, Lawrence HJ, Largman C, Lansdorp PM, et al. Overexpression of HOXB4 in hematopoietic cells causes the selective expansion of more primitive populations *in vitro* and *in vivo*. *Genes Dev* 1995; 9: 1753-1765.
- 91 Antonchuk J, Sauvageau G, Humphries RK. HOXB4-induced expansion of adult hematopoietic stem cells *ex vivo*. *Cell* 2002; 109: 39-45.
- 92 Zhang XB, Schwartz JL, Humphries RK, Kiem HP. Effects of HOXB4 overexpression on *ex vivo* expansion and immortalization of hematopoietic cells from different species. *Stem Cells* 2007; 25: 2074-2081.
- 93 Cheng T. Cell cycle inhibitors in normal and tumor stem cells. *Oncogene* 2004; 23: 7256-7266.